## Segmentation of 3D MicroPET Images Using The Mixture Method

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#### Abstract

The segmentation of 3D microPET image is one of the most important issues in tracing and recognizing the gene activity in vivo. In order to discover and recover the dynamic activity of gene expression, reconstruction techniques with higher precision and fewer artifacts are necessary. To improve the resolution on microPET images, the maximum likelihood estimate (MLE) by the EM algorithm is applied. In addition, advanced statistical technique based on the mixture model is developed to segment the reconstructed images. In this study, the new proposed method is evaluated with simulation and empirical studies. The performance shows that the proposed method is feasibly promising.

*Keywords:* FBP, MLE-EM, Gaussian mixture model, Kernel density estimation, FWHM.

## 1: Introduction

Positron emission tomography (PET) provides a useful medical modality to detect the metabolic activity inside human body. Recent study [1] shows that the technique of microPET has been developed to trace the gene expression *in vivo*. Hence, it is very crucial to enhance reconstruction and analysis techniques with better precision and fewer artifacts so that the genuine gene expression inside biological objects can be recovered.

To improve resolution and reduce artifacts on PET images, many methods have been proposed in literature, including the maximum likelihood estimate by EM (MLE-EM) algorithms and other related techniques. The MLE-EM reconstructions and related improvements have also been proposed [2-12]. The motivation of this study is to apply these techniques with improved analysis methods to reconstruct microPET images.

The filter backpropagation (FBP) reconstruction has been applied in tomography due to its power of fast computation. Wong *et al.* (2001, 2002) use FBP reconstruction and K-means clustering with Akaike information criterion (AIC) to segment PET images. However, the FBP had been developed for transmission tomography. It is not accurate for emission tomography which contains randomness in PET. Moreover, the FBP reconstruction of PET image is typically noisy and inaccurate. Hence, we use the MLE-EM reconstruction to reconstruct microPET images of gene expression more accurately. Due to the varying variances among segments of microPET images, we will consider the Gaussian mixture model (GMM) instead of K-means clustering [16-17].

A rat study by the microPET R4 system in the Institute of Nuclear Energy in Taiwan is used for empirical study. The data consists of 63 slices with 256 by 256 pixels is reconstructed from microPET sinogram. Static scanning of 3600 seconds is applied on rat's brain, the region of interesting (ROI) areas fall between the  $51^{st}$  to the  $60^{th}$  slices.

This study is organized as follows. We present materials and methods in section two. In section three, the evaluated performances of the proposed method are reported by empirical and simulation studies. Finally, we will report our findings and future discussions.

### 2: Materials and Methods

#### 2.1: The MLE-EM Algorithm for PET

Suppose the target image is partitioned into *B* pixels and there are *D* detector tubes. For each pixel, b = 1, 2, ..., *B*, and each tube, d = 1, 2, ..., D, the observations in sinograms of prompt and delayed windows are assumed to follow Eqs. (1) and (2).

$$n_n^*(d) \sim Poisson(\lambda_n^*(d)), \tag{1}$$

$$n_d^*(d) \sim Poisson(\lambda_d^*(d)),$$
 (2)

 $\lambda_{p}^{*}(d) = \lambda^{*}(d) + \lambda_{d}^{*}(d) \qquad \text{and} \qquad$ 

$$\lambda^*(d) = \sum_{b=1}^{B} p(b,d)\lambda(b)$$
. The  $n_p^*(d)$  and  $n_d^*(d)$ 

are the numbers of coincidence events in prompt and delayed windows respectively,  $\lambda(b)$  is the emission intensity of the target image at pixel *b*, p(b,d) is the transition probability,  $\lambda_d^*(d)$  is the accidental coincidence intensity,  $n_p^*(d)$  and  $n_d^*(d)$  are assumed to be statistically independent.

As  $n_p^*(d)$  and  $n_d^*(d)$  are assumed to follow independent Poisson distributions with mean of

where

 $\lambda_p^*(d)$  and  $\lambda_d^*(d)$  respectively, the joint likelihood function of  $L(\lambda_p^*, \lambda_d^*)$  is listed in Eq. (3).

$$L(\lambda_{p}^{*}, \lambda_{d}^{*}) = \prod_{d=1}^{D} \frac{\exp(-\lambda_{p}^{*}(d))\lambda_{p}^{*}(d)^{n_{p}^{*}(d)}}{n_{p}^{*}(d)}$$

$$\prod_{d=1}^{D} \frac{\exp(-\lambda_{d}^{*}(d))\lambda_{d}^{*}(d)^{n_{d}^{*}(d)}}{n_{d}^{*}(d)}.$$
(3)

The EM algorithm can be applied to compute the maximum likelihood estimate iteratively [18-19]. Firstly, the observed data,  $n_p^*(d)$  and  $n_d^*(d)$ , are regarded as incomplete data. The EM algorithm needs to model the complete data. One possible model for PET is shown as in Eqs. (4) and (5).

$$n_p^*(b,d) \sim Poisson\left(p(b,d)\lambda(b)\right),$$
 (4)

$$n_{pd}^{*}(d) \sim Poisson \ (\lambda_{d}^{*}(d)).$$
 (5)

The  $n_p^*(b,d)$  is the detected number of emissions occur at  $b^{\text{th}}$  pixel in  $d^{\text{th}}$  tube and  $n_{pd}^*(d)$  is the detected number of accidental coincidence (AC) events in  $d^{\text{th}}$  tube. It is assumed that  $n_p^*(b,d)$  and  $n_{pd}^*(d)$  are statistically independent, then

$$n_{p}^{*}(d) = \sum_{b=1}^{B} n_{p}^{*}(b,d) + n_{pd}^{*}(d)$$

After applying EM approach, we can determine  $\lambda^{new}$ and  $\lambda_d^{*new}$  as the solutions that maximize the expected log likelihood function in complete data space. The optimal solution can be achieved directly by taking the first derivatives of the new parameters equal to zeros. The solutions are shown in Eqs. (6) and (7).

$$\lambda^{new}(b) = \lambda^{old}(b) \sum_{d=1}^{D} \frac{n_p^*(d)p(b,d)}{\sum_{b'=1}^{B} p(b',d)\lambda^{old}(b') + \lambda_d^{*old}(d)},$$
(6)  
$$\lambda_d^{*_{new}}(d) = \frac{1}{2} \left[ \frac{n_p^*(d)\lambda_d^{*_{old}}(d)}{\sum_{b'=1}^{B} p(b',d)\lambda^{old}(b') + \lambda_d^{*_{old}}(d)} + n_d^*(d) \right],$$
(7)  
for all  $b = 1, 2, ..., B$ , and  $d = 1, 2, ..., D$ .

# 2.2: Gaussian Mixtures Model on 3D Segmentation

For the study of a set of 3D images, it is denoted by  $I_{xyz}$ , where  $I_{xyz}$  represents the intensity at the  $xyz^{th}$  voxel of one set of 3D images. In this study, xy in one slice has the size of 256x256 and the number of slices z has the size of 10. Since the variance scale in segments of 3D images may be different, we consider Gaussian mixture model with different mean vectors and covariance matrices [16-17].

Suppose that the probability of the data  $I_{xyz}$  comes from the  $k^{\text{th}}$  cluster distribution is probability  $\pi_k$ , then, Eq. (8) holds.

$$\sum_{k=1}^{K} \pi_{k} = 1, \text{ and } 0 \le \pi_{k} \le 1, \text{ for all } k = 1, 2, \cdots, K.$$
(8)

Hence, the probability density function of  $I_{xyz}$  is as Eq. (9).

$$f(I_{xyz}; \Phi) = \sum_{k=1}^{K} \pi_k f_k(I_{xyz}; \theta_k),$$
(9)

where  $f_k(I_{xyz}, \theta_k)$  refers to the probability density function in the  $k^{\text{th}}$  cluster with parameter  $\theta_k$ . All parameters are collected to form a parameter vector  $\Phi = (\pi_1, ..., \pi_K, \theta_1, ..., \theta_K)$ . Since, the MLEs are usually difficult to solve directly. Alternatively, the EM algorithm can be applied to find the MLE iteratively [16-17]. Firstly, we introduce an index function as in Eq. (10).

 $C_{xyzk} = \begin{cases} 1, \text{ if } I_{xyz} \text{ comes from } k^{th} \text{ normal distribution.} \\ 0, \text{ otherwise.} \end{cases}$ (10)

Let  $C_{xyz} = (C_{xyzI}, ..., C_{xyzK})$  denote the unobserved index vector and  $\{I_{xyz}, C_{xyz}\}$  form the complete data for the EM algorithm. Given  $C_{xyz}$ , the conditional density of  $I_{xyz}$  becomes Eq. (11).

$$f(I_{xyz} | C_{xyz}) = \sum_{k=1}^{K} C_{xyzk} \pi_k f_k(I_{xyz}; \theta_k).$$
(11)

Similarly using EM approach finds MLE of parameters with the constraint of Eq. (8), the solutions of parameters for the local maximum are shown as in Eqs. (12) - (15).

$$\pi_k^{(new)} = \frac{1}{N} \sum_{xyz=1}^N \omega_{xyzk} , \qquad (12)$$

$$\omega_{xyzk} = \frac{\pi_k^{(old)} f_k(I_{xyz}; \theta_k^{(old)})}{\sum_{i=1}^K \pi_j^{(old)} f_j(I_{xyz}; \theta_j^{(old)})},$$
(13)

$$\sigma_{k}^{2(new)} = \frac{\sum_{xyz=1}^{N} \omega_{xyzk} \cdot (I_{xyz} - \mu_{k}^{(new)})^{2}}{N \cdot \pi_{k}^{(new)}},$$
(14)

$$\mu_k^{(new)} = \frac{\sum_{xyz=1}^{N} \omega_{xyzk} \cdot I_{xyz}}{N \cdot \pi_k^{(new)}}.$$
(15)

Also, the Gaussian distribution for the  $k^{\text{th}}$  cluster is listed as Eq. (16).

$$f_{k}(x;\theta_{k}) = f_{k}(x;\mu_{k},\sigma_{k})$$

$$= (2\pi\sigma_{k}^{2})^{-\frac{1}{2}} \exp\left(-\frac{1}{2}(x-\mu_{k})^{2}/\sigma_{k}^{2}\right).$$
(16)

## 2.3: EM Algorithm for GMM

The EM algorithm for GMM is described as follows.

- 1. Set the initial parameters  $\Phi^{(old)}$ .
- 2. Update the parameters by using Eqs. (12) (15).
- 3. If  $\log L_{in}(\Phi^{(new)}) \log L_{in}(\Phi^{(old)}) < tolerance$ , then the iteration stops. Otherwise, go to Step 2 with the old values of parameters replaced by the new values.

#### 2.4: Slice Normalization

The intensity heterogeneity in slices of microPET images comes from differences in emission activities and reconstruction processes. The heterogeneity between slices affects the accuracy of the segmentation. Hence, slice normalization is needed before 3D segmentation to adjust image levels between slices. The slice normalization is defined in Eq (17).

$$I_{xyz}^{new} = a \frac{I_{xyz} - Min_z}{Max_z - Min_z},$$
(17)

where  $I_{xyz}^{new}$  is the new intensity after slice normalization,  $I_{xyz}$  is the original intensity at  $xyz^{th}$ voxel,  $Min_z$  is the minimum value of the  $z^{th}$  slice,  $Max_z$  is the maximum value of the  $z^{th}$  slice, and a is a none zero constant for adjusted image level, like a =32768.

# 2.5: Determination of the Cluster Number for GMM

The kernel density estimation (KDE) [20] is adopted to determinate the numbers of cluster used in GMM. Those high peaks could be regarded as mean of the groups and low peaks were used to decide standard deviation of the groups. It is based on the empirical rule the range of  $\mu \pm 3\sigma$  covers the most of observations from a normal distribution. Hence, the initialized values of GMM are determined by applying this empirical rule. Let the value of high peak be *hp* and value of the right low peak be *lp*, then this empirical rule is used to estimate  $\sigma$  by (*hp-lp*)/3. Therefore, *hp* and (*hp-lp*)/3 will be used as the starting values of  $\mu$  and  $\sigma$  for one cluster. This approach can automatically decide the cluster number and starting values of each cluster in EM algorithm of GMM.

## **3: Results**

#### 3.1: Simulation Studies

The simulated phantom study with 457932 total counts is displayed in Figure 1. This simulated study is focused on testing and evaluating the performance of GMM. Figure 1A shows target image with five ROIs. Figure 1B displays target image with 50% noise added. Figure 1C presents the clustering results by GMM. The number of clusters is decided by the KDE shown in Figure 1D. There are four local high peaks regarded as the means of four clusters. For accuracy comparison, Figure 2 shows the indices of ROIs by GMM. Figure 3 presents the accurate comparison between the results by K-means and GMM. It is observed that the GMM has a clearer segmentation result than the K-means method

has. Detail results of ROIs are shown in Table 1. The total accuracy of GMM is 92.1% and that of K-means is 66.6%.

Another simulated volume data based on the modified Shepp-Logan's head phantom image is shown as in Figure 4A and 4B. Fifty percentage of noise ratio to phantom images are added. In order to compare the effects of variation between slices, different image levels and shapes of ROIs are considered in slice 1 and 2. First, we use the MLE-EM reconstruction, the result is shown in Figure 4C and 4D. Meanwhile, the GMM is also applied to segment two images without slice normalization as shown in Figure 4E and 4F. It is observed that the boundaries of ROIs are difficult to distinguish. Therefore, slice normalization is applied to the volume data and then GMM is used to segment images as shown in Figure 4G and 4H. The boundaries of these segmentations become clearer after slice normalization. Figure 4I plotted estimated kernel density curve of volume data for finding the number of clusters and initialized values.

For these simulation cases, the performance and accuracy using GMM is better than those of using K-means. The KDE is adopted to decide the number of clusters and the starting values of parameters in the EM algorithm. The slice normalization is necessary when GMM is applied to segment volume data in this study.

### **3.2: Empirical Studies**

The empirical data of one rat (i.e., one big mouse) injected by  $F^{18}$  isotope scanning is collected from the microPET R4 system. The acquired configurations are listed as below. Scanner energy is between 350 and 750 Kev with total scanning 3600 seconds. There are 32 rings in microPET R4 system. File format of histogram data is stored by 2 bytes for each voxel. Ten slices (from the 51<sup>st</sup> to the 60<sup>th</sup> slice) of the volume data are used for investigation and evaluation.

Figure 5 shows estimated kernel density curve of volume data. Based on this KDE, four groups are determinate by local high peaks and their starting values are obtained for EM algorithm. Figure 6 shows the reconstructed rat images by MLE-EM from the  $51^{st}$  to the  $60^{th}$  slice. Besides, Figure 7 and 8 show the segmentation results by GMM and K-means respectively. The detail segmentation from GMM are shown with the comparison to Figure 6. The details uptake areas can be segmented by GMM from  $59^{th}$  and  $60^{th}$  images, in addition, it can segment small areas with high gene expression than those by K-means. On the contrary, the K-means method segments big areas and ignores small uptake areas.

For this real rat microPET study, the GMM leads to more detail segmentation results than the K-means method does. The GMM also has better performance than K-means. The full width half maximum (FWHM) is usually used to evaluate performance of segmented results. The horizontal line profile near the center of the  $60^{\text{th}}$  slice is used to investigate the performance between GMM and K-means. Figure 9 is plotted with four regions in this line profile and their FWHMs for Fig. 6. Table 2 shows that the FWHMs of segmented results by GMM are closer to target FWHMs than those by K-means. Meanwhile, the signal to noise ratio (SNR) defined by the ratio of mean value to standard deviation is used to compare the segmentation performance between GMM and K-means. The SNRs of four regions of GMM are higher than those of K-means.

### 4: Conclusion and Discussion

We have proposed the GMM to segment 3D microPET images with the MLE-EM reconstruction. With fewer artifacts, the MLE-EM reconstruction is more precise than the FBP reconstruction is. For the seeding region growing methods proposed in literature for the segmentation of PET images, initial seeds were crucial to perform images segmentation. The number of clusters was determined by a subjective choice or sequentially searching.

On the other hand, the GMM proposed in this study can perform the segmentation automatically through the KDE method. The KDE approach can automatically estimate the number of clusters and provide the initial values for the EM algorithm of GMM. If the activities of various clusters have different temporary patterns, the slice normalization approach incorporated with GMM is useful to segment 3D images. For further investigation, it will be of great interest to evaluate the qualitative and quantitative performance by more phantom and empirical studies with the comparisons to judgments from medical experts in the future.

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GMM.					
ROI #	True Pixel	Exact Counts		Accuracy (%)	
	Count	K-means	GMM	K-means	GMM
ROI 1	6604	4128	6206	62.5%	94.0%
ROI 2	702	488	522	69.5%	74.4%
ROI 3	748	700	745	93.6%	99.6%
ROI 4	350	280	285	80.0%	81.4%
ROI 5	88	58	59	65.9%	67.0%
Total	8492	5654	7817	66.6%	92.1%

Table 1: Compared clustering results by K-means and

Table 2: The FWHMs and SNRs of segmented results by GMM are better than those by K-means.

Region	FWHM of Region	Pixel of Boundary		Signal to Noise Ratio (SNR)	
		GMM	K-means	GMM	K-means
1	3.75	4	7	9.83	6.82
2	3.40	4	6	8.64	6.23
3	4.14	5	7	4.05	2.13
4	4.45	5	7	3.15	2.13



Fig.1. A) Simulation image of five clusters is displayed. B) Adding 50% noise into Fig. A. C) Clustering results using GMM. D) Kernel density curve using C with values of high and low peaks. It identifies four peaks. Hence, the number of groups is set as four.





Fig.4. Simulated volume data including slice 1 and 2 are marked as A and B. C and D are reconstructed images after added 50% noise ratio to A and B. E and F are segmented results without slice normalization. G and H are segmented results with slice normalization. I is the estimated kernel density curve of simulated volume data after slice normalization.



Fig.5. Kernel density curve using rat volume data (10 slices) with values of high and low peaks. There are four peaks. Hence, the number of groups is set as four. Values of peak are applied to compute starting values in EM algorithm.



Fig.6. The reconstructed rat images are shown from the 51<sup>st</sup> (top-left) to the 60<sup>th</sup> (bottom -right) slice.



Fig.7. The results of segmentation by the GMM are shown from the 51<sup>st</sup> (top-left) to the 60<sup>th</sup> (bottom -right) slice.



Fig.8. The results of segmentation by the K-means method are shown from the  $51^{st}$  (top-left) to the  $60^{th}$  (bottom -right) slice.



Fig.9. The horizontal line profile of the  $60^{\text{th}}$  slice of Fig. 6 is shown with FWHMs. The FWHMs of region 1, 2, 3 and 4 are 3.75, 3.40, 4.14 and 4.45 pixels respectively. The top part shows the location of this line profile in the MLE-EM reconstruction image and the segmented results by GMM and K-means.